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The serologic response of patients with bacillary dysentery (Shigella dysenteriae type 1) was tested by passive hemagglutination, and the results suggest that the test can become a useful adjunct in the diagnosis of this disease. Passive hemagglutination, using cell-wall O polysaccharide from various Shigella, proved to be highly specific for detection of antibodies in rabbit and human sera. Sera from 281 patients with bacillary dysentery had antibodies by the second day after onset of the disease. Maximal titers were reached by the eighth or ninth day. In half of the patients antibody titers fell to nonsignificant levels (< 1:40) by the fifth or sixth month. Antibodies to S. dysenteriae type 1 were sensitive to treatment with 2-mercaptoethanol. On the basis of this sensitivity, their hemagglutinating capacity, early appearance, and rapid catabolism, it appears probable that the antibodies to S. dysenteriae type 1 are in the IgM fraction of serum.

The recent regional epidemic outbreak of Shiga dysentery (Shigella dysenteriae type 1 infection) in Central America provided us with an opportunity to evaluate the specificity of the passive hemagglutination (PHA) test [1, 2] in detecting this infection. Our study describes the pattern of serologic response of patients infected with S. dysenteriae type 1 and contrasts it with that of patients infected with other shigellae.

Materials and Methods

Patients. Three groups of patients were studied: (1) 281 patients with severe clinical dysentery, who were seen from one to 30 days after onset, and most of whom had Shiga dysentery, as determined serologically; (2) 108 patients with

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bacteriologically confirmed bacillary dysentery due to various serotypes of *Shigella*, including the Shiga bacillus; and (3) 43 patients with clinical, bacteriologic, and serologic diagnoses of Shiga bacillus dysentery made three to nine months previously, who could be observed throughout this time span. Some of the cases in this group originally were in the series of 281 patients.

Monospecific sera. Monospecific sera were prepared by the conventional method outlined by Edwards and Ewing [3]. Immune sera were prepared against the epidemic strain of S. dysenteriae type 1, as well as against the serotypes that are commonly isolated in Central America, namely S. dysenteriae type 2, S. flexneri types 1, 3, 4, and 6, and S. sonnei [4–7].

Shigella O polysaccharide. Crude O polysaccharide antigens of Shigella, corresponding to the above serotypes, were prepared by the method of Young et al. [8, 9].

Passive microhemagglutination. Passive microhemagglutination [10] with formalin-preserved erythrocytes [11, 12] was employed. Agreement between duplicate measurements was better than 94% [13]. Antibody titers $\geq 1:40$ were considered indicative of active or recent infection [14].

Treatment with 2-mercaptoethanol. Sensitivity of antibody to 2-mercaptoethanol was determined by incubation of equal volumes of undiluted serum and 0.4 M 2-mercaptoethanol (pH 7.2) for 5 hr at 4 C in the dark. Treated sera were tested in

duplicate and compared with nontreated sera examined on the same day.

Results

Specificity of the hemagglutination reaction. Passive microhemagglutination of each O polysaccharide with monospecific rabbit antisera showed a high specificity for homologous reactions with S. dysenteriae types 1 and 2 and S. sonnei (table 1). The O polysaccharide of other Shigella also reacted, but titers for the homologous system were four- to eightfold higher than for the heterologous system. Similar results were obtained with 108 sera from patients with shigellosis of various serotypes, predominantly S. dysenteriae 1; only one (1.5%) showed significant cross-reaction with the Shiga bacillus (table 2).

Serologic response to S. dysenteriae type 1. In individuals with S. dysenteriae type 1 or 2 whose blood could be examined three to five days after onset and on following days, antibodies were evident promptly after infection and rose to high titers during the first and second week after onset. Of 281 patients with clinically manifest Shiga dysentery who could be examined at various periods after onset, antibodies were demonstrated on the second day (figure 1) in at least one of seven. By the fourth day, six of 15 patients had detectable antibodies. After the ninth day, more than 80% of patients had significant titers of antibody to the Shiga bacillus.

In the group of 43 patients who had had Shiga dysentery within the preceding three to nine months, 80% had positive serologic reactions one month after their illness. Later, there was a progressive decline, so that by the third month, about one-half of the sera had significant titers of anti-

Table 2. Specificity of sera from patients with shigellosis due to *Shigella dysenteriae* type 1 and to other *Shigella*.

Shigella		No. of positive	Significant antibody titer (1:40)			
isolated		patients*	Homologous	Heterologous		
S. dysenteriae	1	70	33 (44.3)†	1 (1.5)		
Other‡		38	16 (42.1)	6 (15.8)		
Total		108	47 (43.5)	7 (6.5)		

- * Confirmed by isolation of the organism.
- † Number of cases with significant titer (percentage).
- ‡ Includes S. dysenteriae type 2, Shigella flexneri types 1 and 6, and Shigella sonnei.

body, and by the ninth month, only 20% had such titers (figure 1).

Titers of antibody to the Shiga bacillus were highest in the first week of the disease (geometric mean titer, 1:480). The mean antibody titer also decreased with time, to a low value after six months (figure 2).

Sensitivity to 2-mercaptoethanol. Sera previously positive showed no significant titers after treatment with 2-mercaptoethanol regardless of the initial value.

Discussion

Hemagglutinating antibodies to the Shiga bacillus could be detected on the first few days of infection, an observation that holds also for other Enterobacteriaceae [15–17]. By the eighth or ninth day, 80% of the patients had significant antibody titers that reached maximal levels after one month. Nevertheless, by the seventh month after onset, few patients had significant levels of antibody. A similar pattern has been reported for other *Shigella* [18, 19].

The humoral immune response to gastrointes-

Table 1. Specificity of HA reactions with antigens and monospecific antisera to Shigella.

	Monospecific antisera								
a	b	С	d	е	f	g			
2,560*	0†	0	0	0	0	0			
0	2,560	0	0	0	0	0			
0	0	10,240	0	0	40	0			
0	0	0	2,560	40	0	0			
0	0	0	160	1,280	0	0			
0	0	160	0	0	10,240	0			
0	0	0	0	0	0	2,560			
	•	•	a b c 2,560* 0† 0 0 2,560 0 0 0 10,240 0 0 0 0 0 0 0 0 0 0 0 0	a b c d 2,560* 0† 0 0 0 2,560 0 0 0 0 10,240 0 0 0 0 2,560 0 0 0 160	a b c d e 2,560* 0† 0 0 0 0 2,560 0 0 0 0 0 0 10,240 0 0 0 0 0 0 0 2,560 40 0 1,280	a b c d e f 2,560* 0† 0 0 0 0 0 2,560 0 0 0 0 0 0 0 10,240 0 0 40 0 0 0 0 2,560 40 0 0 0 0 0 160 1,280 0			

^{*} Numbers are reciprocal titers; boldface numbers indicate homologous reactions.

 $[\]dagger 0$ = Negative in HA test at 1:40 dilution.

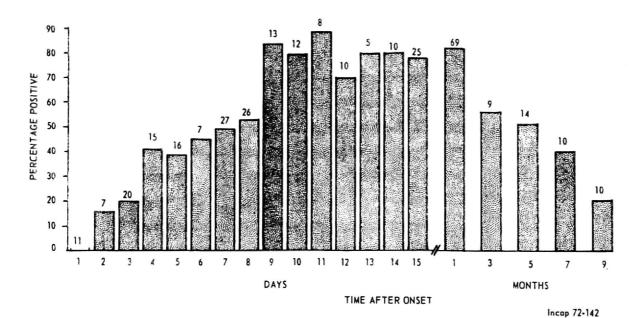


Figure 1. Percentage frequency of patients with significant titers of passive HA antibody to the Shiga bacillus among 281 patients with severe bacillary dysentery studied at various periods in the first two weeks and among 43 patients with proven Shiga dysentery, three to nine months later.

tinal infections depends on the intensity of invasion of the gastrointestinal tract and the extent of damage to the intestinal mucosa [20]. Patients with Shiga bacillus dysentery had higher titers of antibody than patients with bacillary dysentery other than the Shiga type [21, 22]. Antibody titers of patients in our study also were greater than those reported for people vaccinated orally with killed shigellae [23].

Several facts suggest that the antibody we studied belonged in the IgM fraction. Its titer rose quite rapidly, and it declined at a relatively rapid rate; its activity was abolished by treatment with 2-mercaptoethanol. Moreover, its activity was directed against the O polysaccharide, and several studies on experimental animals [24, 25], children

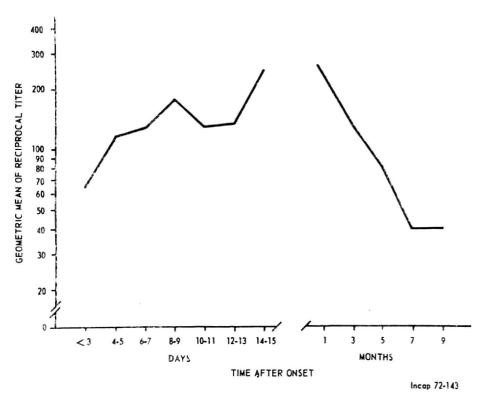


Figure 2. Geometric mean of passive HA antibody titers to Shiga bacillus. The left side of the figure refers to the cases among the series of 281 patients, and the right side refers to the 43 cases observed from three to nine months after onset.

vaccinated with inactivated enteric bacteria [26, 27], and patients with natural infections due to salmonellae [28, 29] have shown that antibodies against enterobacterial O polysaccharide were of the IgM type. Proof of this supposition must await studies by chromatography, ultracentrifugation, and immunochemistry. The passive HA test has proved useful for clinical diagnosis and epidemiologic studies of enteric infections [30-32]. The study showed it to be reliable in the specific diagnosis of Shiga dysentery; 80% of the patients showed significant titers of antibody in the first few days after onset of clinical disease in comparison with similar reported determinations in other shigelloses [21, 32, 33]. Tests repeated several days later confirmed a rise in antibody titers. This serologic test can be a useful adjunct in clinical diagnosis of Shiga bacillus dysentery, but it should not replace isolation of the specific bacillus. On the other hand, this test can be used to great advantage in public health surveillance programs. [34].

The potential usefulness of this test in the diagnosis of dysentery should not becloud the fact that circulating antibodies probably are of little importance in the elimination of infectious agents from the gastrointestinal tract [35–38]. This protection derives mainly from the action of secretory IgA and mechanisms of cellular immunity [39, 43].

References

- 1. Neter, E., Walker, J. Hemagglutination test for specific antibodies in dysentery caused by *Shigella sonnei*. Am. J. Clin. Pathol. 24:1424-1429, 1954.
- 2. Neter, E., Westphal, O., Lüderitz, O., Gorzynski, E. A. The bacterial hemagglutination test for the

- demonstration of antibodies to Enterobacteriaceae. Ann. N.Y. Acad. Sci. 66:141-156, 1956.
- 3. Edwards, P. R., Ewing, W. H. Identification of Enterobacteriaceae. Burgess, Minneapolis, 1962. 258 p.
- 4. Mata, L. Estudio sobre la incidencia de shigelas en Guatemala. Rev. Biol. Trop. 5:211-230, 1957.
- Mata, L. J., Albertazzi, C., Negreros, A., Fernández, R. Prevalence of Shigella, Salmonella and enteropathogenic Escherichia coli in six Mayan villages. Am. J. Public Health 55:1396-1402, 1965.
- 6. Moore, H. A., de la Cruz, E., Vargas-Méndez, O., Pérez, F. I. Diarrheal disease studies in Costa Rica. II. The prevalence of certain enteric organisms and their relationship to diarrhea. Am. J. Public Health 56:442-451, 1966.
- 7. Kourany, M., Vásquez, M. A., Mata, L. J. Prevalence of pathogenic enteric bacteria in children of 31 Panamanian communities. Am. J. Trop. Med. Hyg. 20:608-615, 1971.
- 8. Young, V. M., Sochard, M. R., Gillem, H. C. Infectious agents in infant diarrhea. I. A hemagglutination-inhibition procedure for detection of bacterial fractions in infant sera. Proc. Soc. Exp. Biol. Med. 105:635-638, 1960.
- Young, V. M., Gillem, H. C., Massey, E. D., Baker, H. J. A study on the detection and specificity of antibodies to Shigella flexneri types using preserved polysaccharide-sensitized human erythrocytes. Am. J. Public Health 50:1866-1872, 1960.
- Lee, M. R., Ikari, N. S., Branche, W. C., Jr., Young, V. M. Microtiter bacterial hemagglutination technique for detection of *Shigella* antibodies. J. Bacteriol. 91:463, 1966.
- 11. Beno, D. W., Edwards, E. A. Formalinized red cells in diagnostic virology. Public Health Rep. 81: 377-381, 1966.
- 12. Butler, W. T. Hemagglutination studies with formalinized erythrocytes. Effect of bis-diazo-benzidine and tannic acid treatment on sensitization by soluble antigen. J. Immunol. 90:663-671, 1963.
- 13. Cáceres, A., Mata, L. J. Hemaglutinación indirecta para la investigación de anticuerpos a enterobacteriáceas. Rev. Lat. Am. Microbiol. 12:137–144, 1970.
- Richter, F., Mata, L. J., Cáceres, A. Anticuerpos a Shigella en niños preescolares de Guatemala. Rev. Col. Méd. (Guatemala) 21:85-89, 1970.
- Wentworth, F. H., Brock, D. W., Stulberg, C. S., Page, R. H. Clinical, bacteriological and serological observations of two human volunteers following ingestion of *Escherichia coli* O127:B8. Proc. Soc. Exp. Biol. Med. 91:586-588, 1956.
- 16. Altemeier, W. A. III, Robbins, J. B., Smith, R. T. Quantitative studies of the immunoglobulin sequence in the response of the rabbit to a somatic antigen. J. Exp. Med. 124:443-460, 1966.
- 17. Haltalin, K. C., Matteck, B. M., Nelson, J. D. Microdetermination of shigella hemagglutinins in

- human and rabbit sera with monovalent and polyvalent antigens. J. Immunol. 97:517-524, 1966.
- 18. Neter, E., Dunphy, D. The duration of the hemagglutination response in the serum of children with shigellosis and salmonellosis. Pediatrics 20:78-86, 1957.
- 19. Rauss, K., Pusztai, S., Joó, I., Kétyi, I., Máté, J. Maintenance of artificial dysentery immunity by oral vaccination in humans. Acta Microbiol. Acad. Sci. Hung. 14:153-164, 1967.
- 20. DuPont, H. L., Hornick, R. B., Dawkins, A. T., Snyder, M. J., Formal, S. B. The response of man to virulent *Shigella flexneri* 2a, J. Infect. Dis. 119:296-299, 1969.
- 21. Neter, E., Drislane, A. M., Harris, A. H., Gorzynski, E. A. Study on antibody against enteric pathogens in human gamma globulin. Am. J. Public Health 49:1050-1059, 1959.
- 22. Gotoff, S. P., Lepper, M. H., Fieldler, M. A. Antibody response as an adjunct in the investigation of an outbreak of shigellosis. Am. J. Hyg. 78: 261-268, 1963.
- 23. Yamada, C. An evaluation of shigella vaccines by hemagglutination of human sera from vaccinated volunteers. Jap. J. Med. Sci. Biol. 13:77-90, 1960.
- 24. Fukazawa, Y., Shinoda, T., Tsuchiya, T. Response and specificity of antibodies for Shigella flexneri: demonstration of type-specific factors in immunoglobulin G fraction of antiserum. J. Bacteriol. 98:1128-1134, 1969.
- 25. Tsuchiya, T., Fukazawa, Y., Shinoda, T., Okoshi, T. Response and specificity of anti-shigella rabbit antibody. J. Immunol. 98:1085-1092, 1967.
- 26. Altemeier, W. A. III, Bellanti, J. A., Buescher, E. L. The IgM response of children to Salmonella typhosa vaccine. I. Measurement of anti-typhoid concentrations and their proportions of total serum IgM globulins. J. Immunol. 103:917-923, 1969.
- 27. Altemeier, W. A. III, Bellanti, J. A., Buescher, E. L. The IgM response of children to Salmonella typhosa vaccine. II. Comparison of amounts of IgM specific for the somatic, flagellar and Vi antigens. J. Immunol. 103:924-930, 1969.
- 28. Chernokhvostova, E., Luxemburg, K. I., Starshinova, V., Andreeva, N., German, G. Study on the production of IgG-, IgA- and IgM-antibodies to somatic antigens of Salmonella typhi in humans. Clin. Exp. Immunol. 4:407-421, 1969.
- Wiedermann, D., Wiedermannová, D., Vaerman, J. P., Heremans, J. F. A longitudinal study of serum α₁-antitrypsin, α₂-macroglobulin, transferrin, immunoglobulins IgG, IgA and IgM, and Hand O-agglutinin titers in children following infection with Salmonella enteritidis. J. Infect. Dis. 121:74-77, 1970.
- 30. Young, V. M., Lee, M. R., Branche, W. C., Jr., Kenton, D. M. Shigella flexneri antibody levels in healthy subjects from various regions of the

- United States. Am. J. Public Health 57:2104-2110, 1967.
- 31. Mata, L. J., Cáceres, A., Torres, M. F. Epidemic Shiga dysentery in Central America. Lancet 1:600-601, 1971.
- Mata, L. J., Cáceres, A., Fernández, R., Torres, M. F., Cordón, M., Rosales, R. Avances sobre el conocimiento de la disentería en Guatemala. Rev. Lat. Am. Microbiol. 14:1-10, 1972.
- 33. Neter, E. Epidemiologic and immunologic studies of *Shigella sonnei* dysentery. Am. J. Public Health 52:61-67, 1962.
- 34. Mata, L. J., Gangarosa, E. J., Cáceres, A., Perera, D. R., Mejicanos, M. L. Epidemic Shiga bacillus dysentery in Central America. I. Etiologic investigations in Guatemala, 1969. J. Infect. Dis. 122: 170-180, 1970.
- 35. Honjo, S., Takasaka, M., Fujiwara, T., Kaneko, M., Imaizumi, K., Ogawa, H., Mise, K., Nakamura, A., Nakaya, R. Shigellosis in cynomolgus monkeys (Macaca irus). V. Resistance acquired by the repetition of experimental oral infection with Shigella flexneri 2a and fluctuation of serum agglutinin titer. Jap. J. Med. Sci. Biol. 20:341-348, 1967.
- 36. Tamura, J. T. A cell-free polyvalent shigella vaccine. J. Infect. Dis. 117:353-359, 1967.

- 37. Felsenfeld, O., Greer, W. E., Jiřička, Z. Early immunoglobulin formation and hemagglutinating and bactericidal titers in *Erythrocebus patas* after oral administration of a weakly pathogenic shigella strain. Lab. Invest. 19:146–152, 1969.
- 38. Barksdale, W. L., Ghoda, A. Agglutinating antibodies in serum and feces. J. Immunol. 66:395– 401, 1951.
- 39. Sirisinha, S., Charupatana, C. Antibody response in serum, secretions, and urine of man after parenteral administration of vaccine. Infec. Immun. 2:29-37, 1970.
- 40. Gohar, M. A., Eissa, A. A., Mortada, S. Faecal agglutinins against intestinal pathogens. J. Trop. Med. Hyg. 53:6-9, 1950.
- 41. Gordon, R. S., Bennett, I. L., Jr., Barnes, L. A. Field trial of *Shigella flexneri* III vaccine. III. Coproantibody studies. J. Infect. Dis. 86:197-201, 1950.
- 42. Kenny, J. F., Boesman, M. I., Michaels, R. H. Bacterial and viral coproantibodies in breast-fed infants. Pediatrics 39:202-213, 1967.
- 43. Lodinová, R., Wagner, V. Development of fecal immunoglobulins and coproantibodies in infants after artificial oral colonization with *E. coli* 083. Experientia 26:188, 1970.